

# Pancreas in Preclinical Toxicology

Fifth Conference of Society of Toxicologic Pathology – India

CONTINUING EDUCATION IN TOXICOLOGIC PATHOLOGY -ENDOCRINE AND GASTROINTESTINAL SYSTEM

October 31 - November 2, 2014





#### Introduction



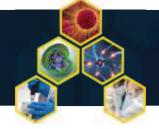
- Only organ in the body with both exocrine and endocrine parts intermixed in parenchyma
  - Islet-Acinar axis
- Finger of the liver- Talmud (200BC-200AD), Cushion of Stomach (Vesalius)
  - Wirsung-pancreatic ducts (1642), de Graaf-secretions (1664), Digestive actions- 1834-1844
  - Enzymes-Trypsin, amylase, lipase- 1879, enterokinase- 1889, Histology-Langerhans-1869
  - Disease first reported only in 19<sup>th</sup> century, >500 drugs affecting pancreas



#### Introduction

- Silent and specialized organ
  - Not involved in absorption of ingested xenobiotics
  - Not involved in detoxification
  - Synthesize and secrete variety of specific proteins
- Low frequency of xenobiotic induced toxicity
  - When toxicity occurs, major impact on health
- Toxicity is more common where exposure to environmental pollutants, contaminants and alcohol is high
- Organ is pale with grossly visible lobules and ducts but not islets, except in prominent peri-insular holos
- Compact in hamster, primates, dogs and humans with comparable anatomy, less compact in rodents, diffuse in rabbits





#### **Exocrine Pancreas**

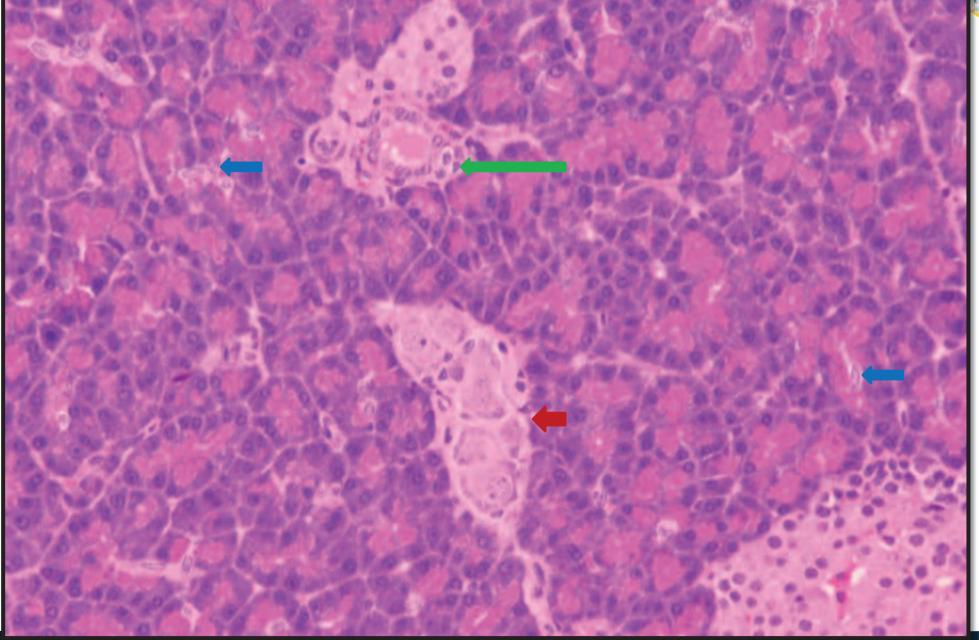


- ~90% consists of acinar cells and ductal cells, centroacinar cells
- Acinar cells, pyramid shaped, convoluted, often interconnecting, tubuloacinar network in lobules, cyclic changes in morphology
- Basophilic cytoplasm around nucleus due to RER, apical granular eosinophilia due to zymogen granules digestive enzymes
- Pancreatic stellate cells –play major role in disease and cancer
- Centroacinar cells- interface b/n acinar cells & intercalated duct, due to their tonofibrillary architectures, act as a valve regulating the amount of secreted zymogen granules
- Intercalated duct continues as Intralobular ducts basement membrane of extracellular matrix



# Pancreas Histology





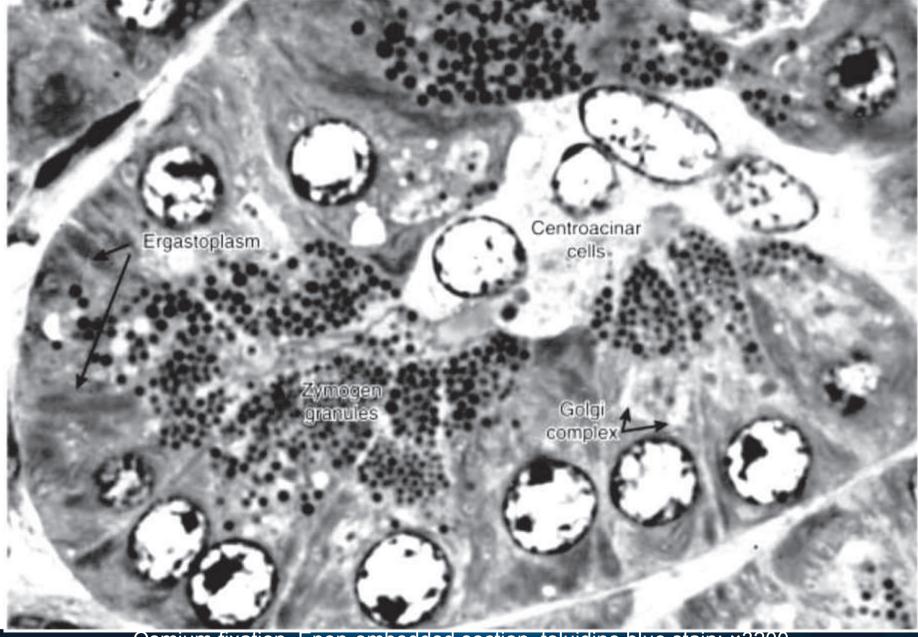
#### **Exocrine Pancreas**

- Ductal epithelium- cuboidal or low columnar
- Interlobular ducts collagenous layer
- Exocrine pancreatic secretion is tightly regulated by neuroendocrine system
- Species specific differences in physiology pathobiology of pancreatic disease in animal models relevance to human disease
- Rats and mice- gastric lobe, duodenal head and tail
- Peri-insular and tele-insular regions, greater in mice than rats
- Acinal and ductal cells can divide in experimental animalsregeneration can occur



#### **Electron Microscopy**





Osmium fixation. Epon-embedded section, toluidine blue stain; ×3200. (From Bloom W, Fawcett DWA: Textbook of Histology, 11th ed. Philadelphia, WB Saunders, 1986)<sup>7</sup>

# Species differences in distribution of drug metabolizing enzymes in pancreas



TABLE 1.—The distribution of CYPs and GSTs in the ductal cells of the syrian hamster (SGH), nude mouse (NM), rat, rabbit, guinea pig, dog, monkey, and human.

СҮР	SGH	NM	Rat	Rabbit	Pig	Dog	Monkey	Human
1A1	+	_	+	+	_	++	+	+
1A2	+	—	_	_	—	—	_	+
2B6	+	++	+	_	—	—	+	-/+a
2C8,9,19	+	++	+	_	_	_	+++	++b
2D1	+	+	+	++	+	+	+	++
2E1	_	+	+	+	_	_	_	-c
3A1	+	++	++	++	+	++	++	++
3A2	÷	+	+	+	++	+	++	+
3A4	÷	++	_	_	+	_	_	++
$GST-\pi$	++	+	+	_	÷	++	+++	++
GST-α	+	<u> </u>	++	_	÷	++	++	++
GST-μ*	++	++	+	++	+	++	+++	++ <sup>*,d</sup>

Staining intensities: -, none; +, weak; ++ moderate; +++ strong.

TABLE 3.—The distribution of CYPs and GSTs in the islet cells of the syrian hamster (SGH), nude mouse (NM), rat, rabbit, guinea pig, dog, monkey, and human.

CYP	SGH	NM	Rat	Rabbit	Pig	Dog	Monkey	Human
1A1	++	+	++	+++ <sup>a</sup>	+++	+	++	+/+++ <sup>b</sup>
1A2	+++	+++ <sup>a</sup>	$+^a$	+++ ++ <sup>a</sup>		-		+ _/+_+_b,d
2B6 2C8,9,19	+++ <sup>a</sup> +	+++	+++a	++	+++ <sup>b</sup>	_	+++-	_/+++ <sup>b,e</sup>
2D1	÷	+++	++	+	+	+	÷	+++f
2E1	+++	++	+++		-	b	. —	$-/+^{h}_{h}$
3A1 3A2	+++ <sup>a</sup>	++	+++	++	— —	++ <sup>b</sup>	+++	++"
3A4	+	++	+++	_	_	<u> </u>	+	<b>_</b> /+++ <sup>k</sup>
GST- $\pi$	+++	$++^a$	+	+	_	++	++	$-/+++^{m}$
$GST-\alpha$	$+++^a$	+	-	_		-		<b>-</b> /+ <sup>n</sup>
GST-μ*	++	+	+	$+++^{a}$	$+^{b}$	-	++	++*

TABLE 2.—The distribution of CYPs and GSTs in the acinar cells of the syrian hamster (SGH), nude mouse (NM), rat, rabbit, guinea pig, dog, monkey, and human.

СҮР	SGH	NM	Rat	Rabbit	Pig	Dog	Monkey	Human
1A1	++	_	_	+	_	++	+	+
1A2	+	-	++	-	—	+++	+	+
2B6	+	++	_	_	+	++	++	$-/+^{a}$
2C8,9,19	++	++	_	_	++	+	++	$++^{b}$
2D1	++	+++	_	_	+	+	+	++c
2E1	_	+	+	+	_	_	_	$-/+^{d}$
3A1	++	++	+	++	+	++	++	+
3A2	++	+	+	++	++	++	_	+
3A4	+	+	+	+	++	_	++	+
GST-π	_	_	<u> </u>	_	_	_	_	<u> </u>
GST-α	_	_	_	_	++	_	+	$++^{e}$
GST-μ*	_	_	_	_	_	_	_	+*

Staining intensities: -, none; +, weak; ++ moderate; +++ strong.

- Most enzymes expressed in human, hamster, mouse, and monkey.
- Several isozymes are lacking in rats, pigs, rabbits, and dogs
- Islet cells of all 8 species express more enzymes than ductal and acinar cells

#### **Pancreatitis**

- Types of pancreatitis
  - Toxic Alcohol(2), pesticides, xenobiotics
  - Obstructive- gallstones(1), tumors, worms, congenital defects
  - Metabolic- hyperlipidemia, hyperkalemia, acidosis
  - Infectious- parasitic, viral, bacterial
  - Genetic- cystic fibrosis transmembrane conductance regulator, pancreatic secretary trypsin inhibitor 1, serine protease inhibitor Kazaltype 1, cationic trypsinogen 1
  - exocrine pancreas has great reserve capacity. No clinical evidence of exocrine pancreatic insufficiency up to 90% destruction
- Sorafenib, mesalazine, azathioprine, simvastatin definite pancreatitis inducing drugs; alcohol, corticosteroids, estrogens, opiates.
- dasatinib, nilotinib, and bosutinib- up to 70% increase in serum amylase activity and/or lipase activity with no frank pancreatitis



## **Animal models of Pancreatitis**

- Invasive Models of Pancreatitis
- Noninvasive Models of Pancreatitis
  - caerulein-induced pancreatitis
    - abnormal secretory vesicle maturation & formation of large cytoplasmic vacuoles
    - acinar cell injury with both autophagy and apoptosis observed as end points (and occasionally necrosis)
    - cholecystokinin antagonists effective, proglumide and benzotript
  - muscuranic receptor agonist- carbachol, OP- fenthion & diazonin
    - augmenting the normal pancreatic flow and increase the intrapancreatic ductal pressure



## **Animal models of Pancreatitis**

- Alcohol-induced Pancreatitis
  - diets rich in unsaturated fat, with gradually ascending ethanol administration regime - reproducible, chronic alcohol-induced pancreatitis
- Choline deficient diet containing 0.5% DL-ethionine
- Genetic Animal Models of Pancreatitis
  - CFTR Deficient Tg Mouse, SPINK3 KO Mouse, PRSS1 overexpressing Transgenic Mouse Model
- Amylase, Lipase and pancreas-enriched miRs-216a and -375, promise as novel biomarkers for pancreatic injury



#### **Exocrine Pancreas**



#### **INHAND** nomenclature and diagnostic critiera

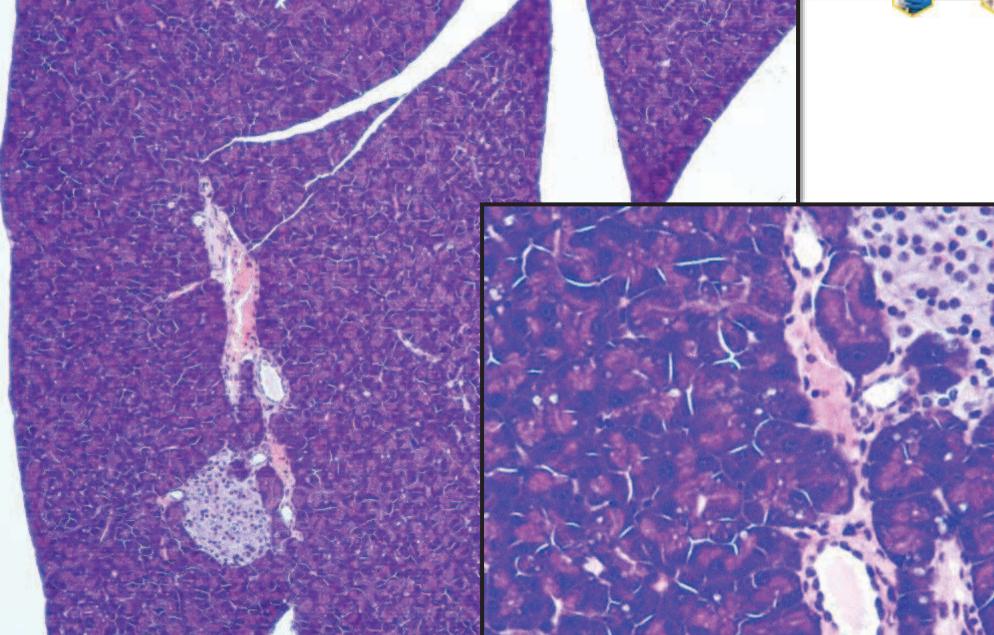
- a. Congenital Lesions
  - Ectopic tissue
- b. Cellular Degeneration, Injury and Death
  - Vacuolation, acinar cell
  - Fatty infiltrate
  - Eosinophilic globules
  - Autophagic vacuoles, acinar cell
  - Single cell necrosis/apoptosis
  - Necrosis
  - Zymogen granules decreased, acinar cell
  - Atrophy, acinar cell
  - Mineralization
  - Amyloid
  - Pigment
- c. Inflammatory Lesions
  - Infiltrate
  - Inflammation

- d. Vascular Lesions
  - Inflammation, vessel
  - Hemorrhage
- e. Miscellaneous Lesions
  - Hypertrophy, acinar cell
  - Halos, peri-insular
  - Focus, basophilic
  - Metaplasia, hepatocytic
  - Metaplasia, ductular
  - Ectasia, duct
  - Edema
  - Fibrosis
- f. Non-neoplastic Proliferative Lesions
  - Hyperplasia, acinar cell
  - Hyperplasia, ductal cell
- g. Neoplasms
  - Adenoma, acinar cell
  - Adenoma, ductal cell
  - Adenocarcinoma, acinar cell
  - Adenocarcinoma, ductal cell



# VACUOLATION, ACINAR CELL

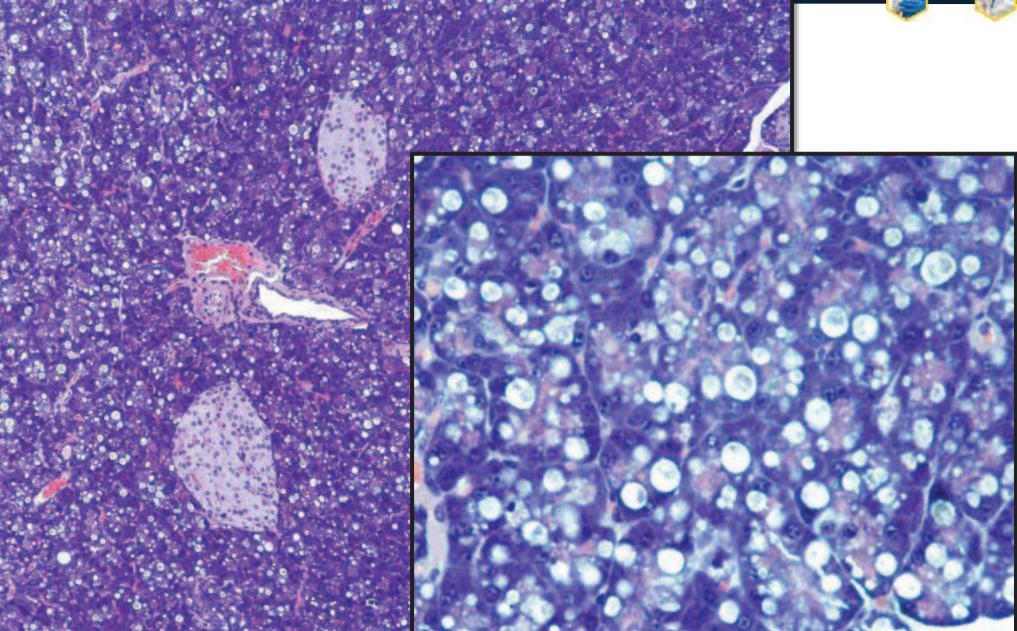




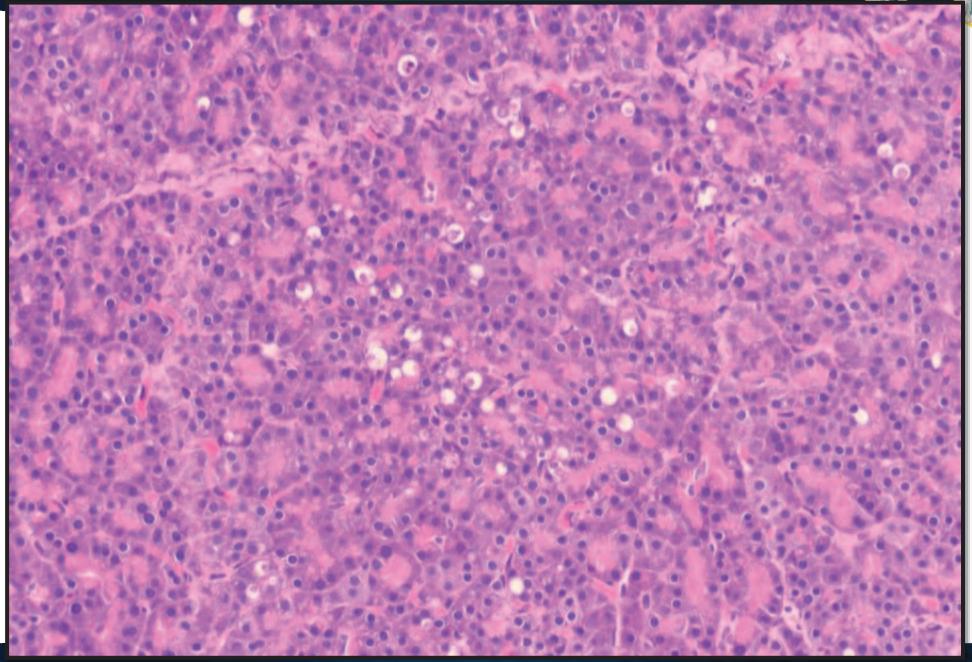


## VACUOLATION, ACINAR CELL

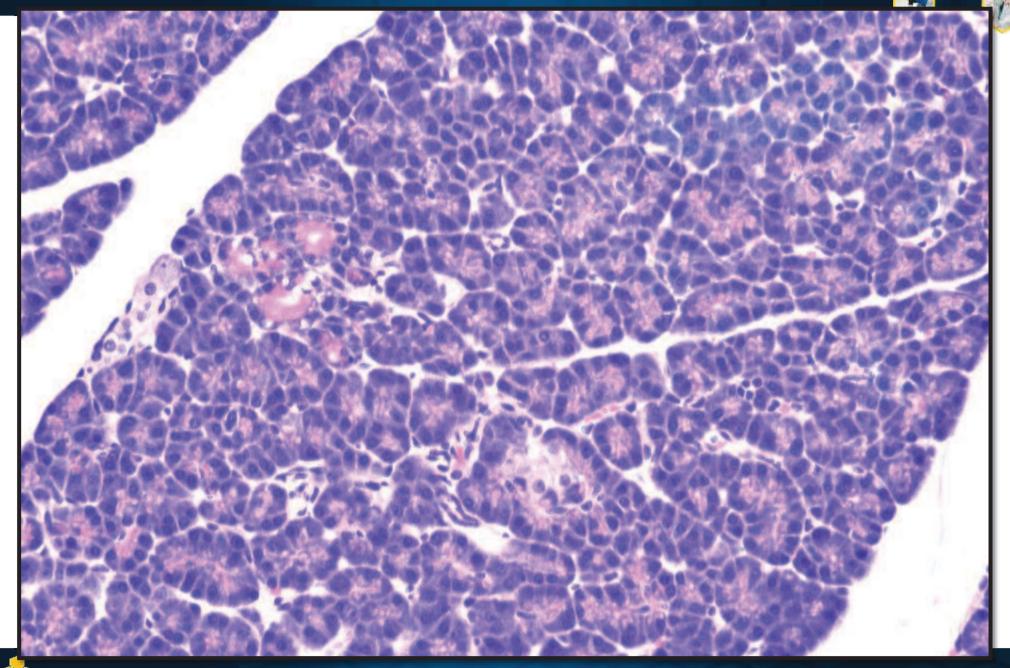




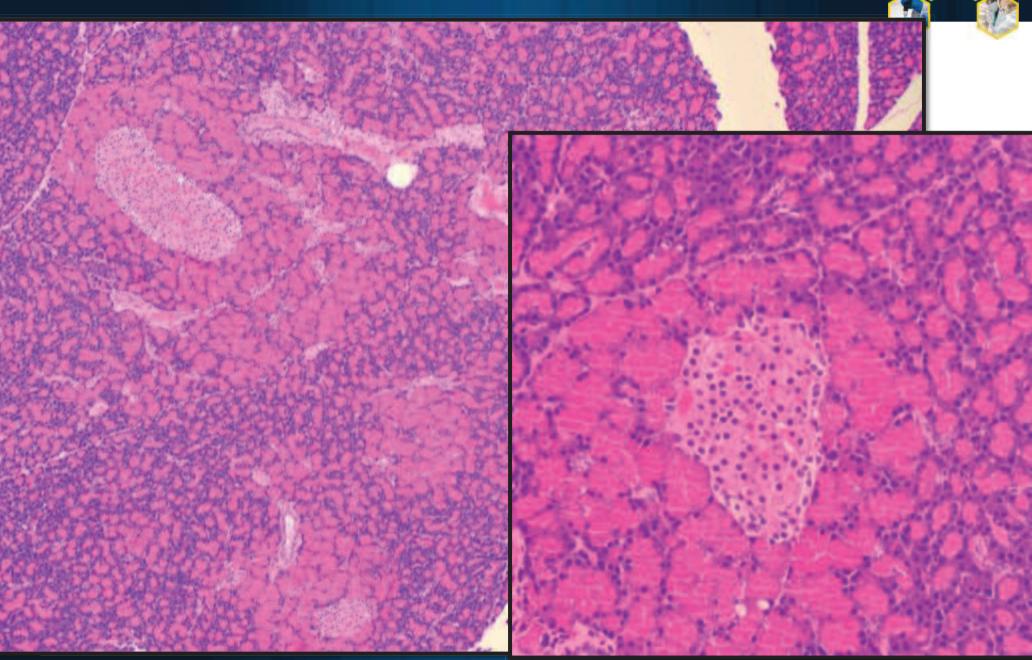
## **AUTOPHAGIC VACUOLES, ACINAR CELL**



## **ZYMOGEN GRANULES DECREASED, ACINAR CELL**

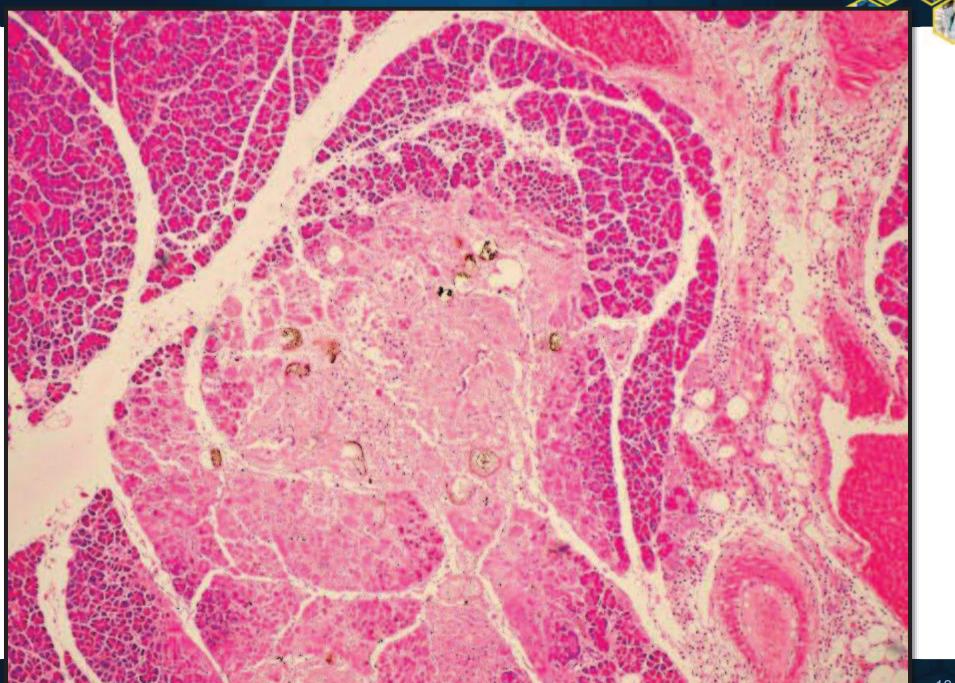


# HALOS, PERI-INSULAR

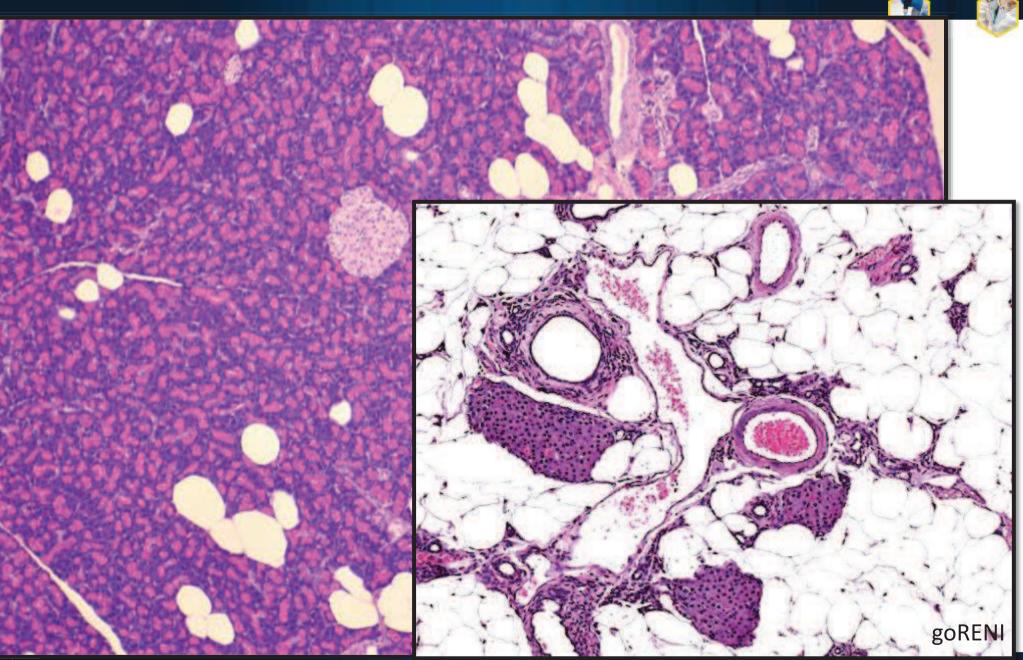




## NECROSIS



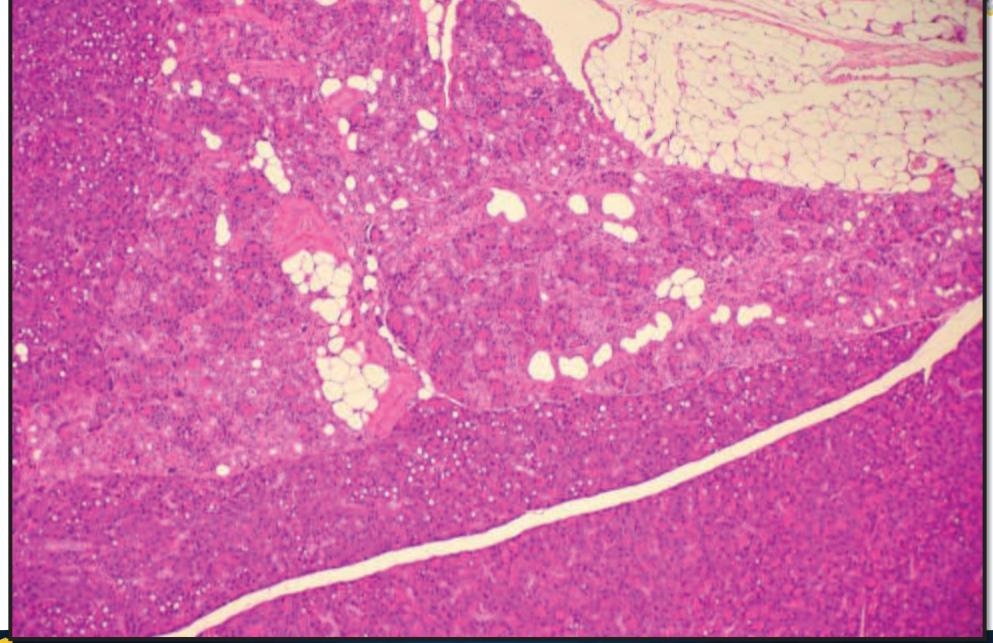
## FATTY INFILTRATE



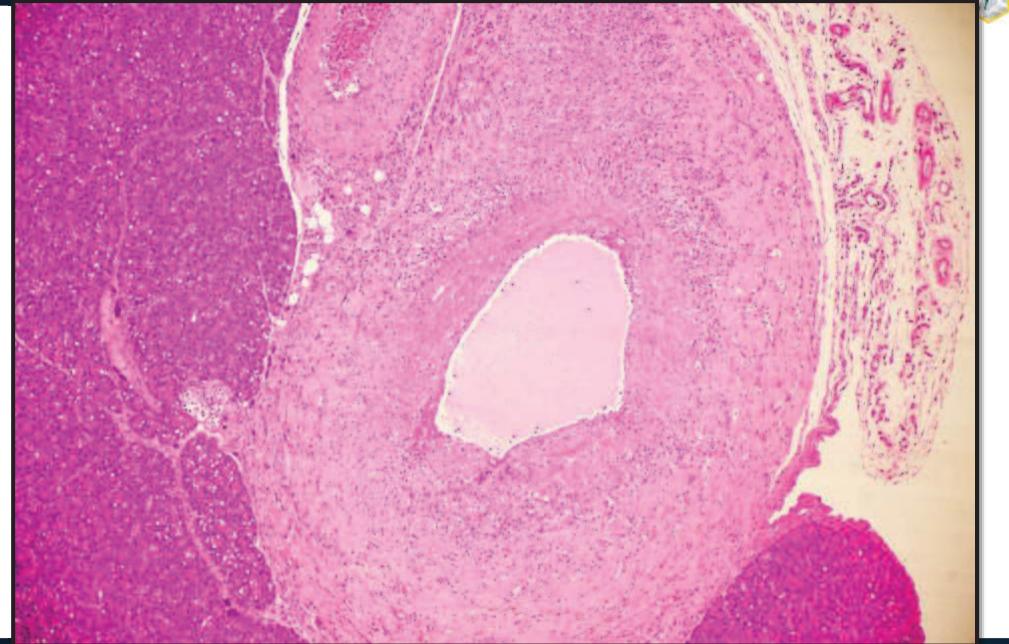


# ATROPHY, ACINAR CELL





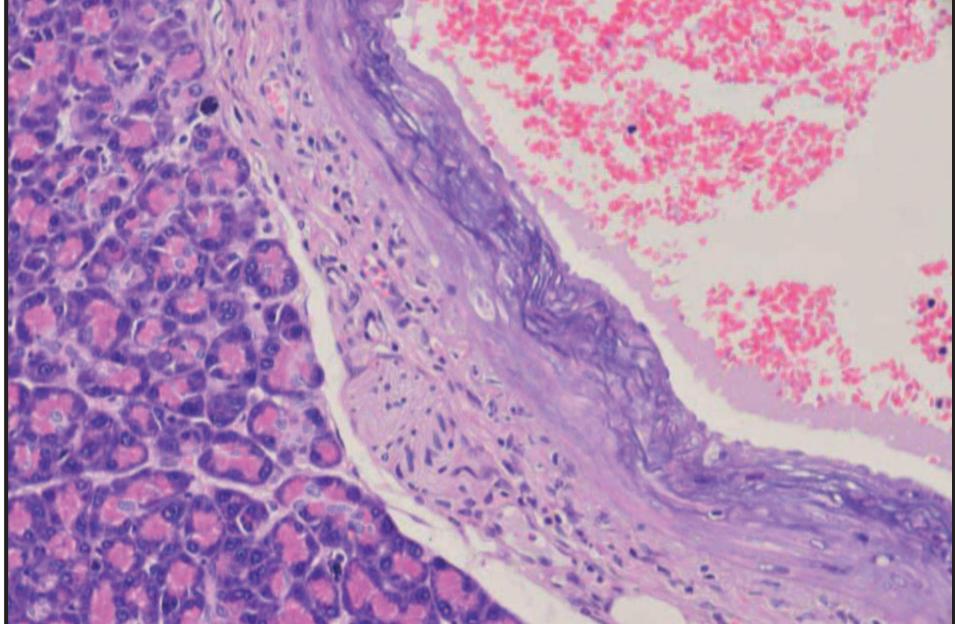
## **INFLAMMATION & HYPERTROPHY, VESSEL**





## Mineralization- blood vessels

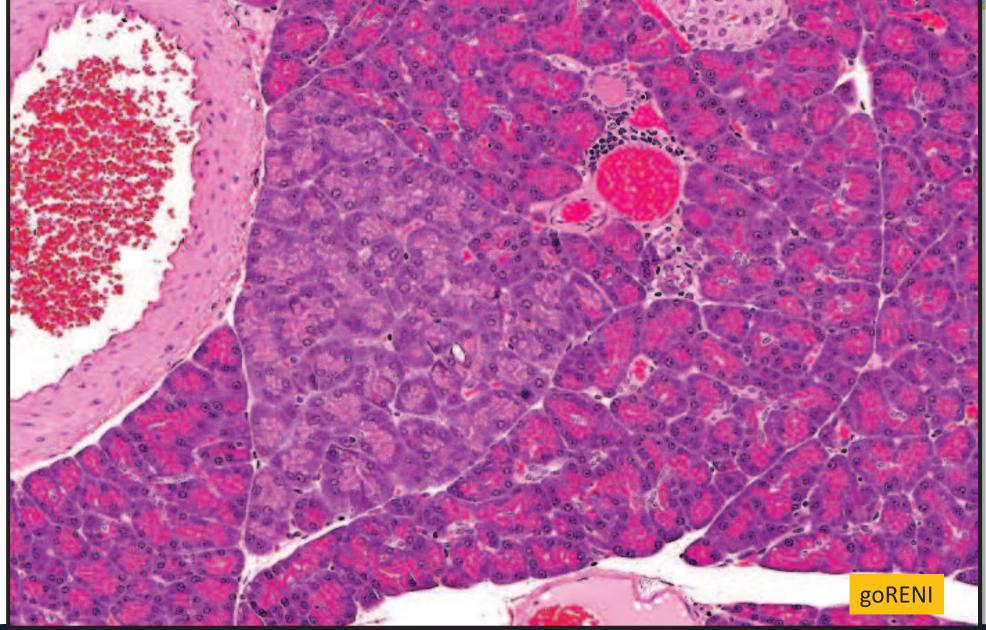






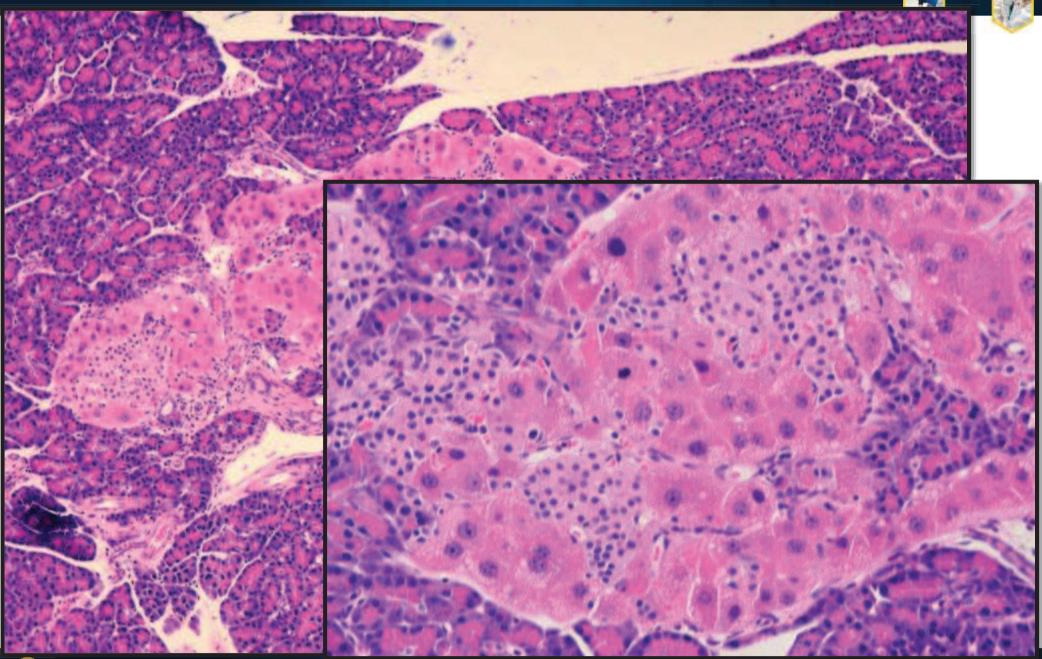
# FOCUS, BASOPHILIC





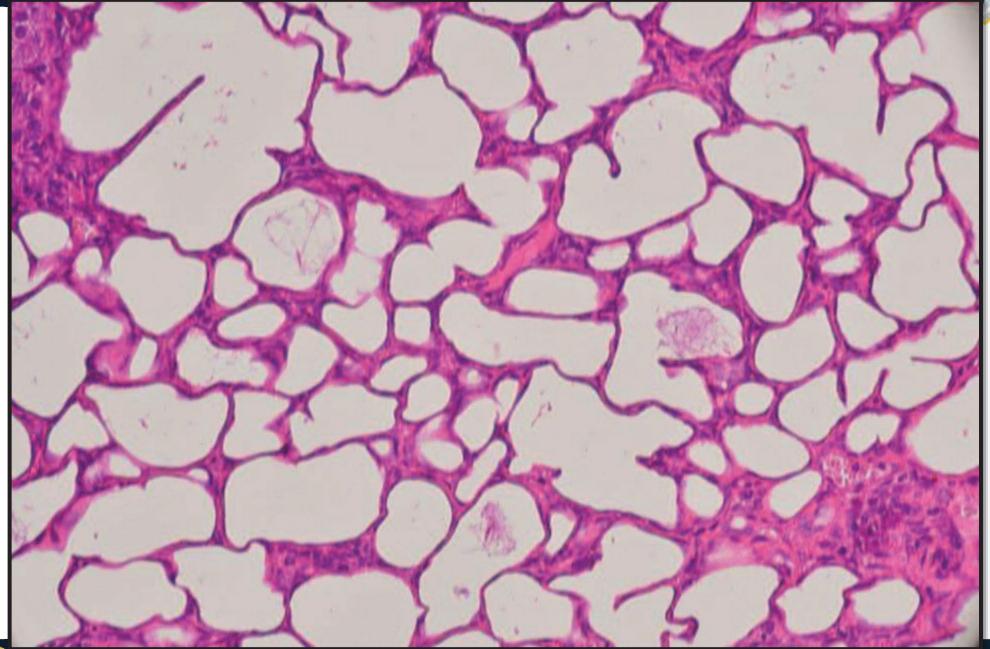


# **METAPLASIA, HEPATOCYTIC**

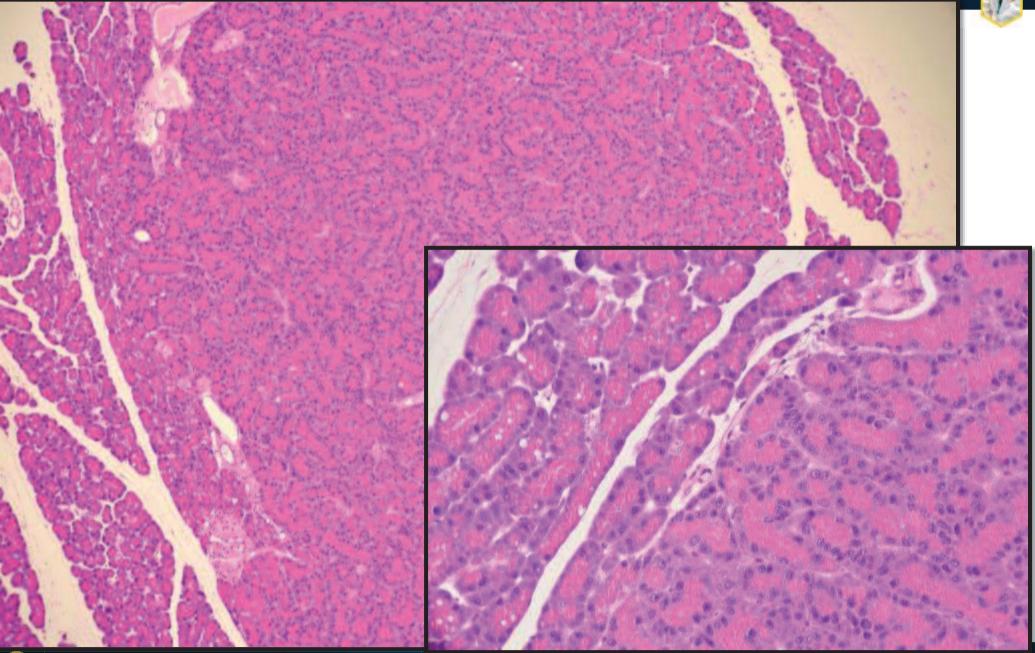




# ECTASIA, DUCT

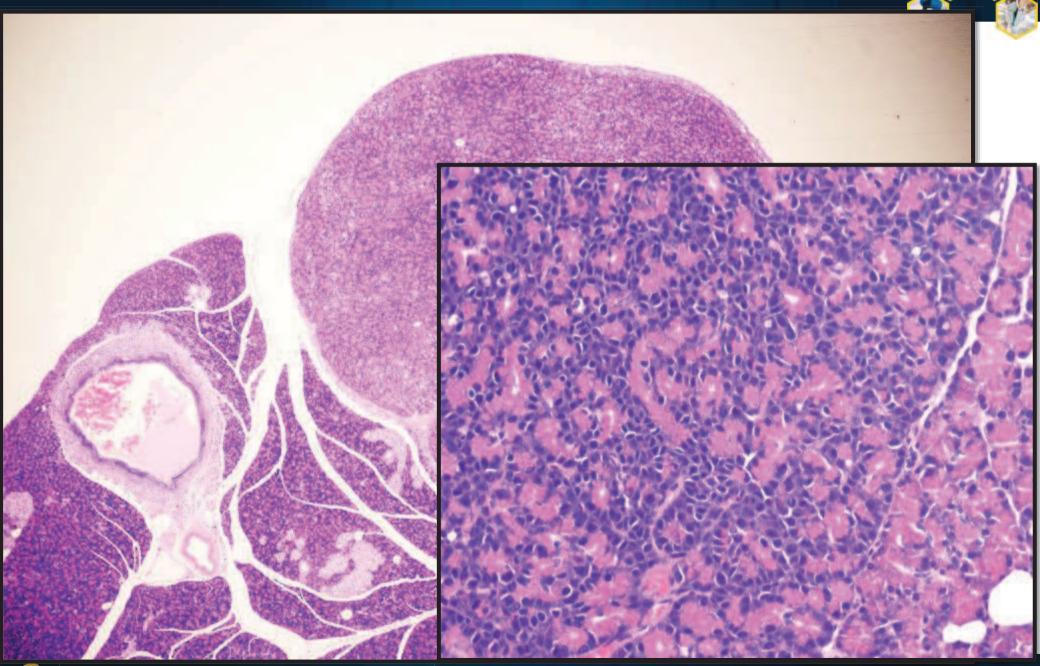


# HYPERPLASIA, ACINAR CELL





# ADENOMA, ACINAR CELL





#### **Endocrine Pancreas**

- 1-2% of parenchyma receives 20% of blood supply
- Capillary network in islets is 10 time more fenestrated
- Several islet derived hormones and peptides influence directly exocrine function
- α cells glucagon, β cells insulin & amylin, δ cells somatostatin, ε cells ghrelin, PP or F cells- PP & adrenomedullin
  - Stimulatory PACAP, NO, VIP, A II
  - Inhibitory SST, PP. ghrelin, pancreastatin, adrenomedullin, galanin, CGRP, NPY and PYY



#### **Endocrine Pancreas**



#### **INHAND** nomenclature and diagnostic critiera

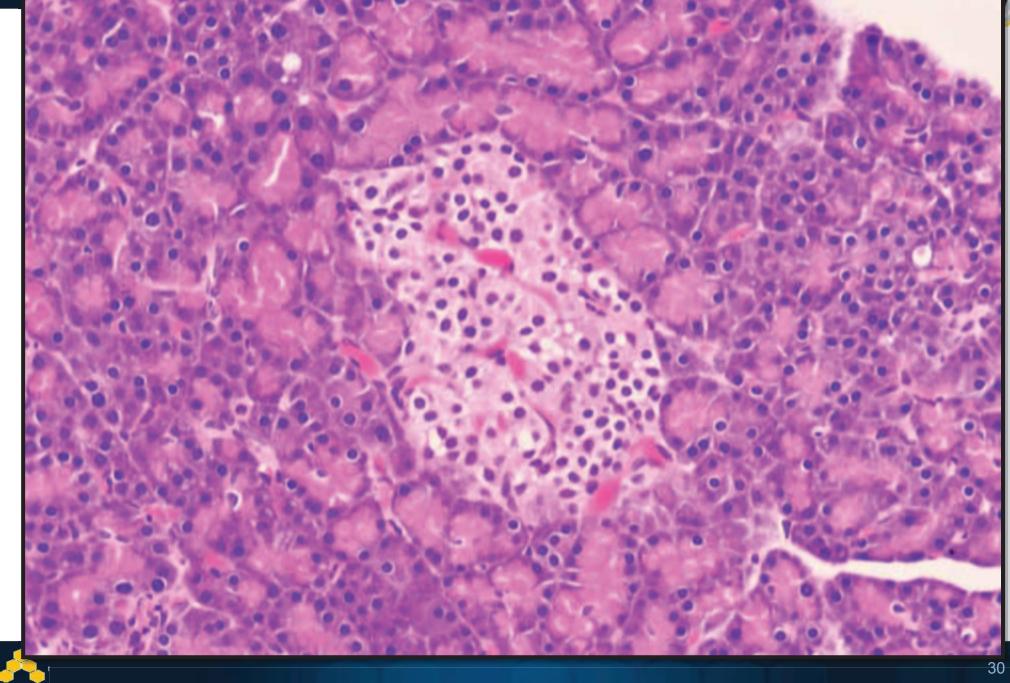
- Non proliferative
  - Amyloidosis, islet
  - Angiectasis
  - Apoptosis, islet
  - Atrophy, islet
  - Degranulation, ß-cell
  - Fibrosis, islet
  - Infiltrate, inflammatory cell
  - Inflammation
  - Metaplasia, hepatocyte
  - Necrosis, single cell
  - Pigmentation, islet
  - Vacuolation, ß-cell

- Proliferative
  - Hyperplasia, islet cell
  - Adenoma, acinar-islet cell
  - Adenoma, islet cell
  - Carcinoma, islet cell



# VACUOLATION, ß-CELL





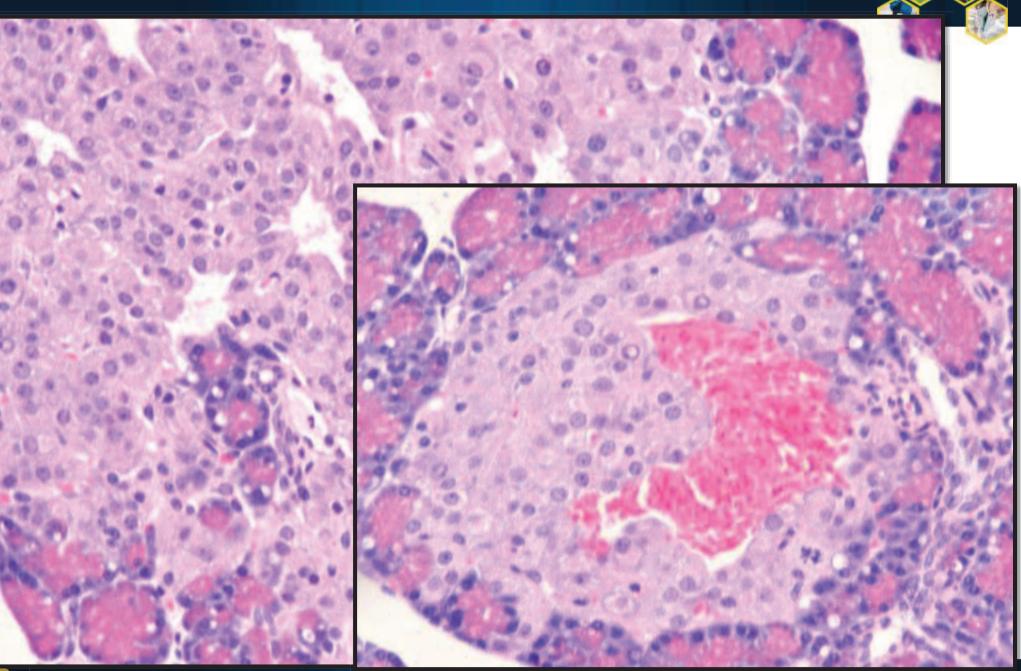
## **AMYLOIDOSIS, ISLET**





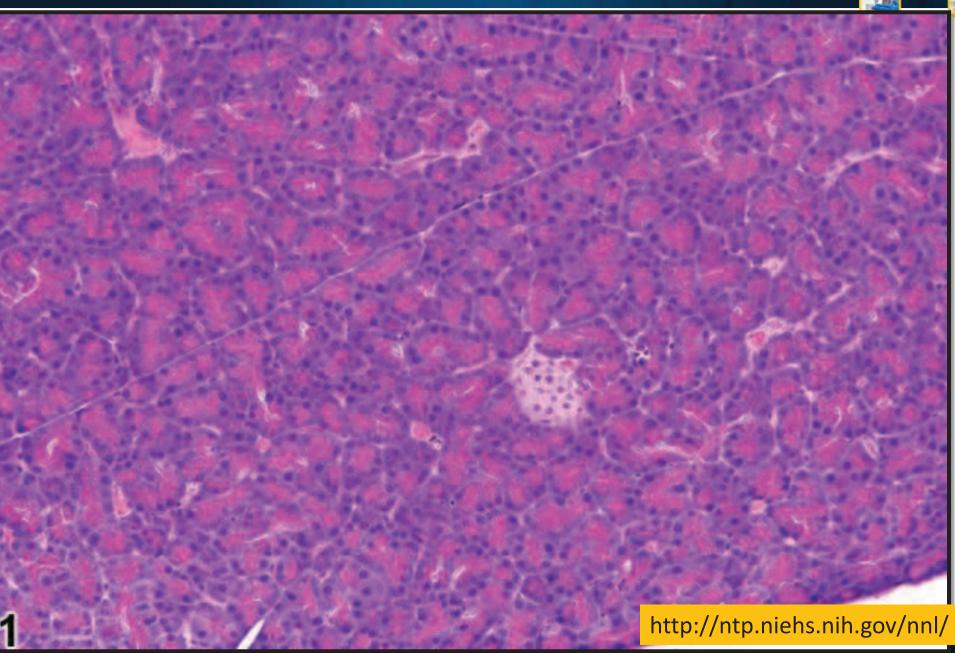
*Toxicol Pathol* 1991 19: 123

# ANGIECTASIS



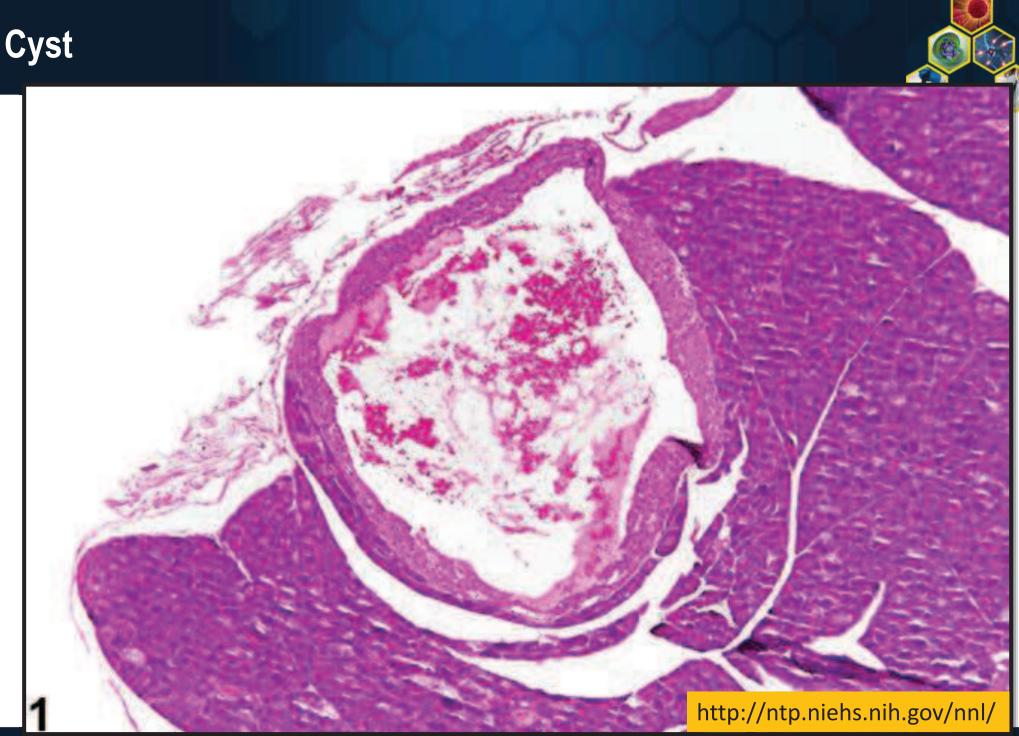


## ATROPHY, ISLET

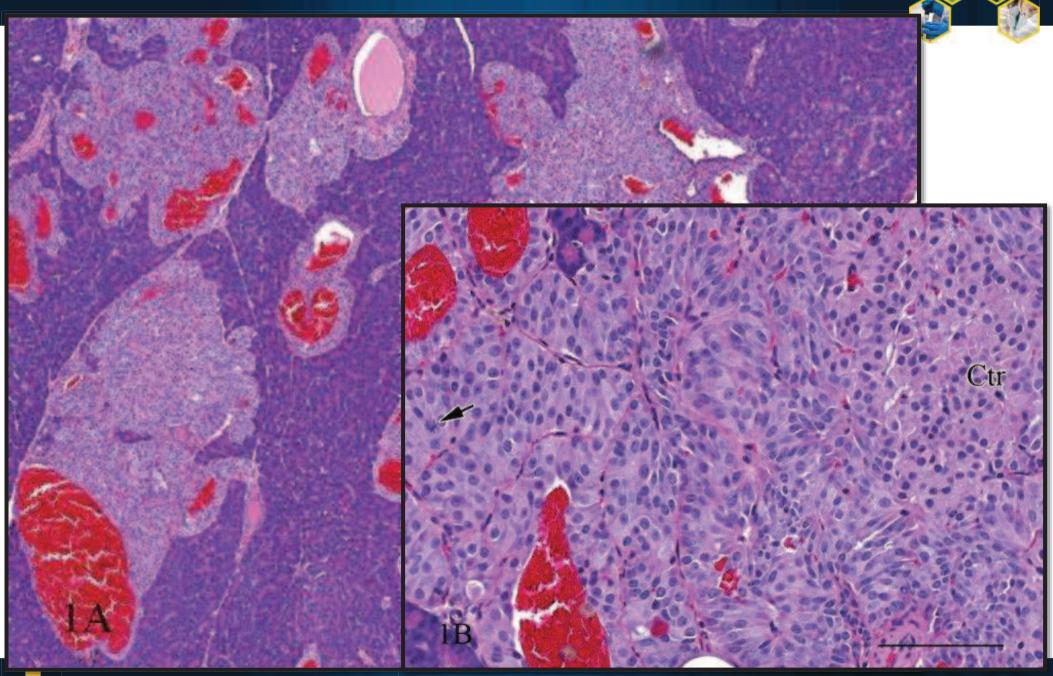








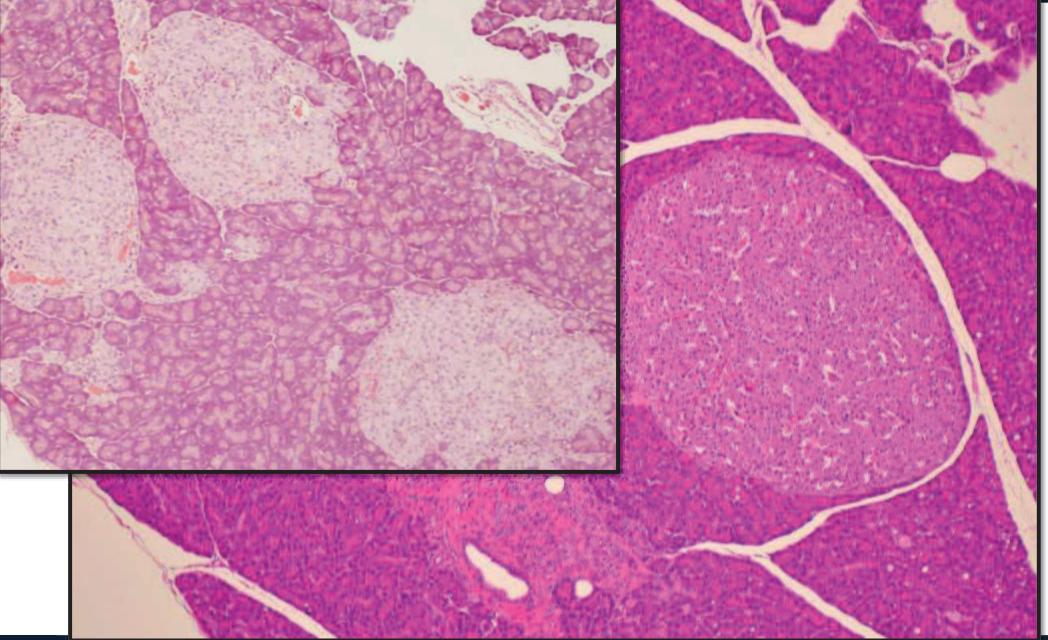
## **Atypical Case of Islet Cell Hyperplasia**





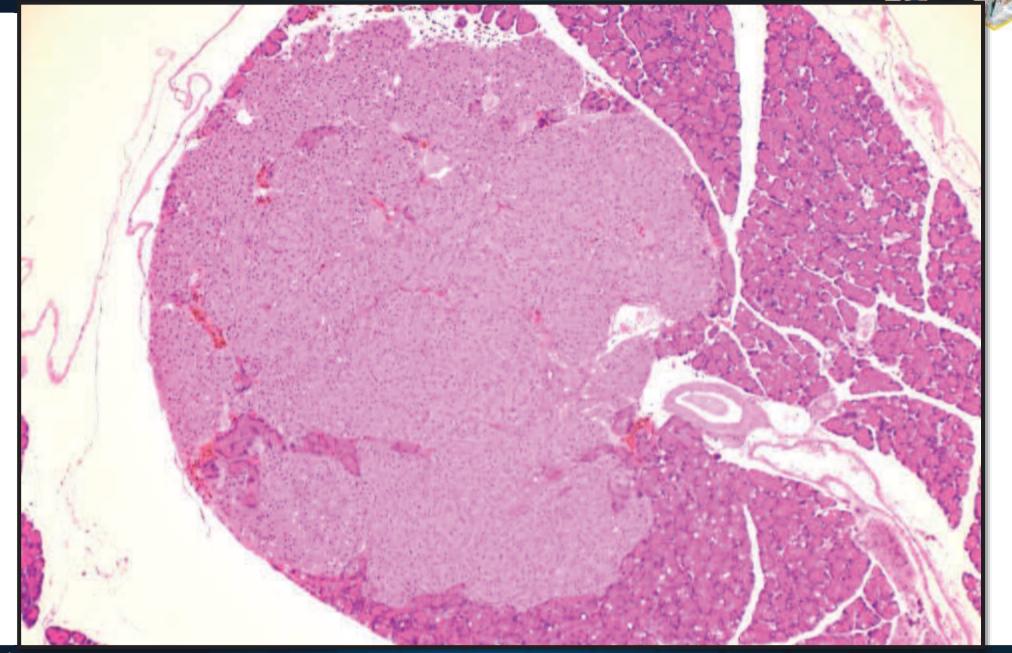
# HYPERPLASIA, ISLET CELL



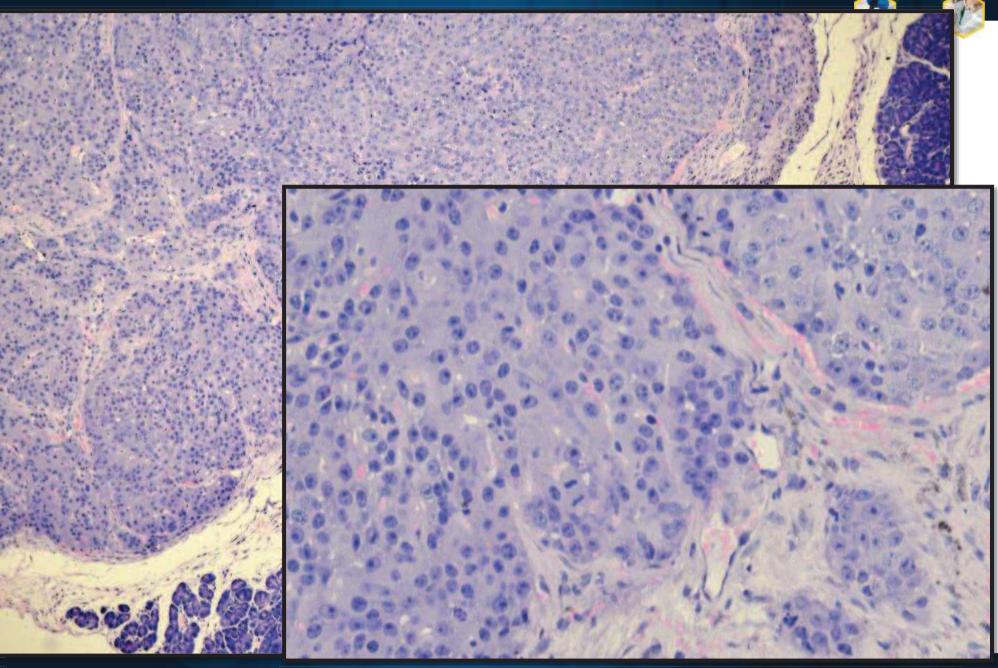




# ADENOMA, ISLET CELL

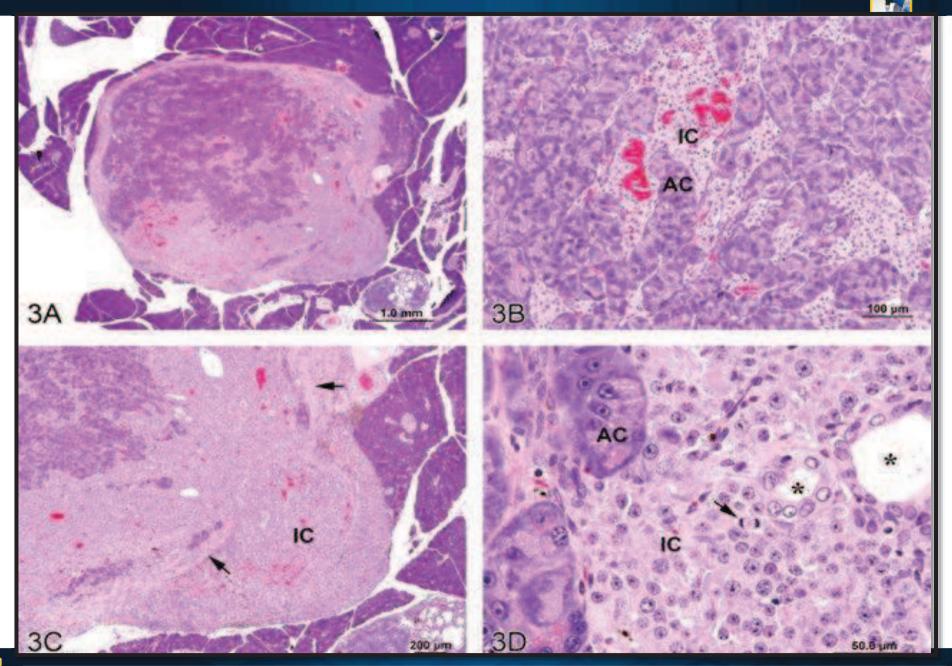


# CARCINOMA, ISLET CELL





## ADENOMA, ACINAR-ISLET CELL



Adams E T et al. Toxicol Pathol 2010;39:240-266

## ACKNOWLEDGEMENTS







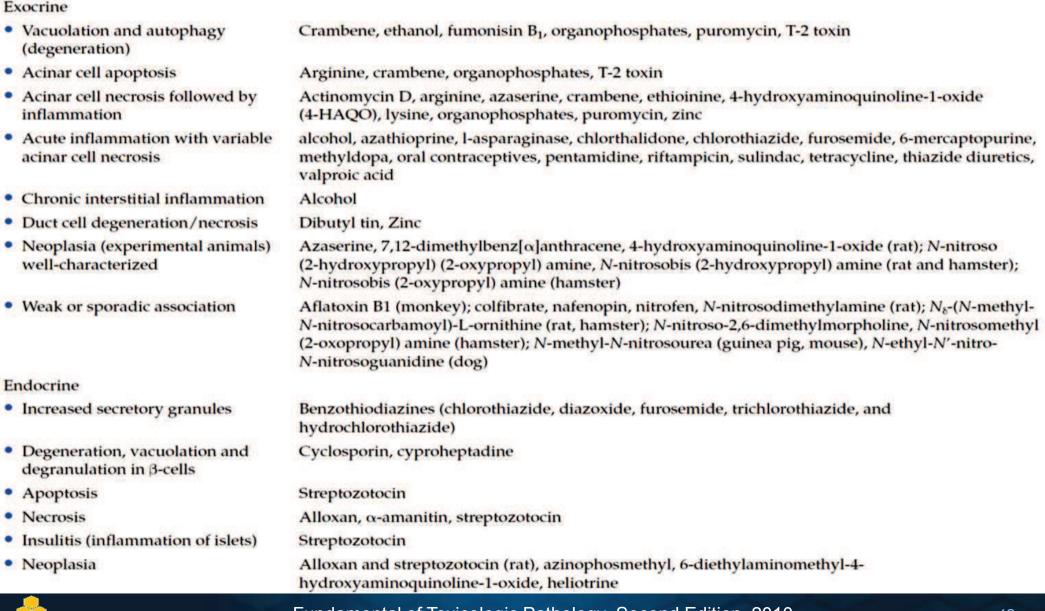
# Thanks

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#### Classification of pancreatic alterations with selected examples



Xenobiotic





Fundamental of Toxicologic Pathology, Second Edition, 2010